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20 4 BIOPHYSICAL MECHANISMS

21 This chapter considers mechanisms of interaction between radiofrequency (RF) electromagnetic fields
22 (EMFs) and biological systems. Such an approach is necessary in order to fully understand any biological effects
23 that have been observed and to extrapolate effects observed under specific experimental conditions to understand
24 what might occur in other situations. In the absence of an understood biophysical mechanism, the plausibility of
25 any reported biological effects is reduced. The chapter draws principally upon the work contained in international
26 expert group reports from the International Agency for Research on Cancer (IARC, 2013) and the International
27 Commission on Non-Ionizing Radiation Protection (ICNIRP, 2009). A quantitative review of potential
28 mechanisms of interaction between RF fields and biological systems by Sheppard et al. (2008) is also of particular
29 note.

30 It is well understood that exogenous (externally arising) RF fields can penetrate into the body tissues
31 and deposit the energy they carry as heat (see Chapter 3). Moreover, irrespective of the initial mechanism of
32 interaction involved, all energy deposited in biological tissue is ultimately transformed to heat. Health-related
33 guidelines (ICNIRP, 1998) and standards (IEEE, 2005) have been developed based on heating effects and with the
34 objective of restricting temperature rises in the body to levels where no harm is expected to result.

35 Biological effects that occur as a result of heating are generally termed *thermal effects*, whereas those
36 that occur through mechanisms other than heating are termed *non-thermal* effects. However, it should be
37 recognised that effects that do not occur as a result of heating could still occur in the presence of significant
38 heating and that this broad categorisation is not always helpful.

39 As an over-arching principle applying to biophysical interactions, Challis (2005) explains that
40 biological systems (like any other system) are subjected to random fluctuating electric and magnetic fields known
41 as thermal noise. Therefore, in order for a system to respond to an applied RF field, the size of fields induced in
42 the system should be larger than the corresponding random fields.

43 Sheppard et al. (2008) categorised biophysical mechanisms as either *established* or *proposed*. The
44 established mechanisms relate to plausible biological effects which have been proven to occur through rigorous
45 experiments. On the other hand, the proposed (unproven or not established) mechanisms are those that have not
46 yet gained acceptance through rigorous experiments yielding consistent results, and often have been developed to
47 explain particular experimental observations.

48 Sheppard et al. (2008) concluded that the dominant established mechanisms are dielectric relaxation¹
49 and resistive loss which lead to energy deposition and increase of tissue temperature through heating. Pearl chain
50 formation in uniform electric fields (see section 4.4.7), nonlinearity and generation of harmonic frequencies, and
51 demodulation are also amongst established mechanisms operating over all or parts of the RF range. Electron
52 tunnelling and the radical pair mechanism were examples of proposed (unproven) mechanisms. Most other non-
53 thermal mechanisms considered were based upon coupling to specific vibrational modes in molecules, cells or
54 tissues. These mechanisms are not discussed here in detail and would generally be excluded from health-related
55 discussions because, to be biologically effective, they would be accompanied by temperature rises that would
56 overwhelm any other biological response.

57 4.1 Ionization potential of RF fields

58 Electromagnetic radiation comprises energy that is carried in quanta known as photons. The energy of
59 each photon is proportional to its frequency, and the total energy carried by the radiation depends on the number of
60 photons and the energy carried by the individual photons. The photon energy, E (in joules), for a radiation of a
61 frequency, f (in hertz), is given by:

$$62 \quad E = hf$$

¹ Dielectric relaxation relates to the process through which electrical charges that have become displaced on an object such as a cell move back to their equilibrium positions after the externally applied field that caused them to become displaced is removed. This is an exponential process governed by a characteristic of the material known as the relaxation time. The relaxation time is related to frictional forces that slow the movement of charges and it results in a delay in the movement of charges in response to an imposed time-varying field. The frictional forces and delay in the movement of charges lead to energy loss.

63 where h is Planck's constant, equal to 6.626×10^{-34} Js and equivalent to 4.136×10^{-15} eVs, noting that 1 eV equals
64 1.602×10^{-19} J.

65 At radio frequencies the energy of a photon varies from 4.14×10^{-10} eV at 100 kHz to 4.14×10^{-6} eV at a
66 typical telecommunications frequency of 1 GHz and to 1.24×10^{-3} eV at 300 GHz. This is much smaller than the
67 energy required for ionization by ejection or promotion of orbital electrons from atoms of the material through
68 which an electromagnetic wave propagates. The precise threshold energy for ionization depends on the type and
69 state of matter. For example, the minimum photon energies required for removing an electron from biological
70 molecules such as calcium, glucose and water are 6.1, 8.8 and 11.2 eV respectively (Bushberg et al., 2011). Since
71 water constitutes the most abundant molecular target for ionization in living organisms, 11 eV is often taken as the
72 lower limit for ionization in biological systems.

73 As demonstrated above, a single photon of RF radiation has relatively low energy; therefore, it is not
74 capable of causing ionization. This is why electromagnetic radiation in the RF spectrum is regarded as non-
75 ionizing radiation. Ionizing radiations such as gamma- and X-rays have photon energies above 11 eV and so their
76 damaging effects largely result from ionization taking place in biological cells and tissues. Such effects are not
77 produced by a single photon of RF radiation (Lin, 1978).

78 In addition to removal of an electron from atoms or molecules, photons can interact with materials by
79 breaking chemical bonds, if they have sufficient energy. The energy required to break various bonds that are found
80 in biological systems has been quantified (Masamichi, 2006). Typical covalent bonds require 1–10 eV, and typical
81 hydrogen bonds require 0.1 eV. Thus, even at a frequency of 300 GHz, the photon energy of RF radiation is still
82 two orders of magnitude too small to break hydrogen bonds, which are the weakest form of chemical bond.

83 The possibility of adverse effects due to multiple photon absorption at the same site was studied by
84 Prohofsky (2004). In principle, multiple photon processes could shift the incident energy to a region where
85 resonant interactions are known to occur. However, multi-photon absorption requires very intense incident photon
86 beams so the probability is raised that a sufficient number of photons will interact simultaneously with an electron
87 to deliver the amount of energy needed for excitation. Pickard and Moros (2001) demonstrated that this probability
88 is very low, even under an extreme scenario with an SAR of 100 W/kg, which exceeds restrictions in exposure
89 guidelines (ICNIRP, 1998) and is likely to heat the tissue by several degrees.

90 **4.2 Classical interaction mechanism (heating)**

91 As mentioned above, the most recognised and well established mechanism through which biological
92 effects of RF radiation can occur is tissue heating through dielectric and resistive energy losses (Sheppard,
93 Swicord & Balzano, 2008).

94 The absorption of RF EMF energy by biological systems generates oscillating motions of charged
95 particles and water molecules, which are strongly dipolar and are the major component of biological tissues. Polar
96 molecules move to align themselves with the EMF to minimize their potential energy. Motion and resonant
97 oscillations in polar subgroups of macromolecules (e.g. proteins, DNA) are largely damped by collisions with
98 surrounding water molecules. The damping imposes strict limitations on the energy that can be accumulated in
99 vibrational or rotational modes. If the modes are over-damped (i.e. with decay times less than the time-period of
100 the RF oscillations), resonant absorption does not occur. Damping precludes a direct RF pumping of such modes
101 (Sheppard, Swicord & Balzano, 2008). These collisions disperse the energy of the RF signal into random
102 molecular motion. Tissue heating occurs because the rotational motion of molecular dipoles (including those of
103 water itself) is hindered by the viscosity of water and interactions with other molecules, i.e. the rotational energy is
104 transferred to the surrounding aqueous environment as heat.

105 In the absence of strong damping by water molecules, compact molecules have their lowest intra-
106 molecular vibrational modes at frequencies no lower than approximately 200 GHz. Acoustic modes² can exist in
107 long flexible molecules at arbitrarily low frequencies. However, over-damping by water precludes energy
108 absorption by all modes for frequencies up to the far infrared. Moreover, molecular absorption spectroscopy is
109 usually performed at frequencies above 300 GHz because the spectrum of substances below this frequency usually

² *Acoustic modes* occur when adjacent molecules in a lattice move in the same direction as each other during an oscillation whereas *optical modes* occur when adjacent molecules move successively towards and apart from each other during the oscillation.

110 presents as a continuum without the defined peaks in absorption at certain frequencies that would be characteristic
111 of resonant vibrational motion (Prohofsky, 2004).

112 The magnitude of motion that results from the interaction of polar substances with electric fields
113 depends on the strength and frequency of the field. In addition, the actual increase in temperature depends on the
114 ability of the organism to thermo-regulate. At higher radiofrequencies, above a few GHz, where the orientation of
115 dipoles cannot keep up with the oscillations of the field, the system behaves like a non-polar substance (Stuchly,
116 1979).

117 The energy deposition also depends on the dielectric properties of the cells or tissue, and the local field
118 properties. In principle, it would appear possible to create very high spatial gradients for heat generation within
119 tissue, either because of localised micro-field structure, or because of local enhancements of dielectric property.
120 Nevertheless, even if such conditions were to occur, thermal diffusion in tissue prevents the creation of high
121 spatial gradients in temperature such that localised hot-spots cannot occur on the cellular scale (Liu & Cleary,
122 1995).

123 Standards for RF exposure of people are based on protection against adverse effects that might occur
124 due to increases in tissue temperature (head and spinal cord to 38 °C, neck and trunk to 39 °C and limbs to 40 °C).
125 For testes, which are normally at a temperature somewhat below the normal 37 °C core body temperature, the
126 increase in temperature is limited to 1 °C (NRPB, 2004b). An increase in core body temperature of 1°C or less
127 corresponds to a whole body SAR of ~4 W/kg (ICNIRP, 1998). Because RF energy penetration and induced
128 effects are dependent on the frequency of the incident field and the composition of exposed tissues, quantifying
129 SARs in small averaging regions is also relevant for evaluations of localised heating and/or any other effects. For
130 frequencies up to a few GHz, as used in wireless communications, SAR is normally averaged over either 1 or
131 10 g. Hirata et al. (2008) suggested that the chosen averaging mass should be that which maximises the correlation
132 with local temperature elevation. The dominant factors influencing the correlation between mass-averaged SAR
133 and temperature elevation are the thermal diffusion length in the biological tissue, which largely depends on the
134 blood perfusion rate, and the penetration depth of the RF waves (Hirata, Ito & Fujiwara, 2009).

135 In a typical exposure scenario where an individual is exposed to RF radiation during the use of a mobile
136 phone against the side of the head, the RF-generated temperature rise in the brain, ranges from 0.05 to 0.12 °C per
137 W/kg (NRPB, 2004a). It is unlikely that any biological effect in the brain would be caused by these small
138 increases in temperature (Repacholi, 2001).

139 Temperature changes approaching 1 °C are likely to affect several biological processes, although some
140 temperature-sensitive molecular and physiological effects may occur with smaller increase of temperature, i.e.
141 ≤ 0.1 °C (Foster & Glaser, 2007). Rates of temperature increase can also be important in causing a physiological
142 change. Microwave-induced hearing has been attributed to a rapid rate of heating of head tissue, 1–10 °C/s, which
143 leads to acoustic waves being formed due to expansion of tissue water. This auditory effect is associated with brief
144 pulses (1–10 μ s) at frequencies of 1–10 GHz and peak power-densities of $\sim 10^4$ W/m² and occurs with only small
145 increases in temperature in the head (Foster & Glaser, 2007).

146 **4.3 Induced charge and dipole relaxation**

147 An external RF field can translate and rotate charged and polar molecular structures as well as other
148 cellular components of biological materials. The magnitude of these motions depends on the strength and
149 frequency of the field and may be impeded by inertia and viscous forces. The orientation of polar molecules under
150 the influence of external fields does not occur instantaneously and follows a time-dependent behaviour known as
151 the relaxation process. It also takes some time after the application of an external field for electric charges within
152 the cells and tissue structures to accumulate at the interfaces and reach a new equilibrium state (relaxation).
153 Depending on the size/characteristics of polar molecules, different types of relaxation processes can take place in
154 biological tissues.

155 In isolation, small charged particles, such as monopolar ions, are able to respond at frequencies up to at
156 least 10^{12} Hz (in the infrared), but the association of ions with water molecules (solvation) means that the dielectric
157 properties of water, with its large dipole moment, are dominant in biological solutions. Water molecules can rotate
158 freely in an oscillating low frequency electric field with little energy loss; however, at frequencies above 10^8 Hz,
159 the rotational inertia of the molecules begins to inhibit rotation, causing energy absorbed from the field to be
160 dissipated by collisions or nearest neighbour interactions in the medium medium (Sheppard, Swicord & Balzano,
161 2008).

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162 When a dipole distribution is uniform, the positive charges of one dipole cancel the effect of the
 163 negative charges from another adjacent dipole. However, when the dipole distribution varies from point to point, a
 164 complete cancellation cannot occur. Therefore, an uncancelled charge would be left at an interface surface, which
 165 becomes an equivalent bound charge in the material. The relaxation process may therefore be illustrated by
 166 considering the response of bound charges to an applied electric field (Lin, 2000; Michaelson & Lin, 1987). In this
 167 case, the dynamic force balance equation is given by

$$168 \quad m \frac{d^2 x}{dt^2} = qE - m\omega_s^2 x - m\nu \frac{dx}{dt} \quad (4.1)$$

169 where E is the applied electric field, x is the displacement of a charged particle in the direction parallel to the field,
 170 ω_s is the characteristic frequency of the elastic, spring-mass system, ν is the particle collision frequency, and m and
 171 q are the mass and charge of the particle, respectively. The force exerted on the particle, mass multiplied by
 172 particle acceleration on the left-hand side of equation 4.1, results from an electric driving force qE , an elastic
 173 restoring force in proportion to displacement, x , with elastic constant denoted as $m\omega_s^2$, and a retarding damping
 174 force proportional to velocity, dx/dt , with damping coefficient, $m\nu$.

175 After Fourier transformation and rearranging terms, equation (4.1) becomes

$$176 \quad x(\omega) = [(q/m)E]/[\omega_s^2 - \omega^2 + j\omega\nu] \quad (4.2)$$

177 where ω is the angular frequency of the applied field and equal to $2\pi f$. Note that the equilibrium position for the
 178 charge ($x = 0$) represents local charge neutrality within the medium. When the charge is displaced from its
 179 equilibrium position, a dipole is established between the charge itself and the “hole” that is left behind and bound
 180 in the molecular and membrane structure. A dipole moment p is formed by the charge q times the displacement x .
 181 For a medium with volume-bound charge density ρ , the total dipole moment per unit volume or polarization P is

$$182 \quad P = \rho p = [\rho(q^2/m)E]/[\omega_s^2 - \omega^2 + j\omega\nu] \quad (4.3)$$

183 The electric flux density D may be expressed in terms of the electric field E and polarization P as

$$184 \quad D = \epsilon_0 E + P \quad (4.4)$$

185 where ϵ_0 is the vacuum or free space permittivity. For isotropic media, the permittivity may be related to D by the
 186 expression $D = \epsilon E$. These relations together with equation (4.3) give an equation for the permittivity,

$$187 \quad \epsilon(\omega) = \epsilon_0 [1 + (\omega_p^2)/(\omega_s^2 - \omega^2 + j\omega\nu)] \quad (4.5)$$

188 where

$$189 \quad \omega_p^2 = \rho q^2 / m\epsilon_0 \quad (4.6)$$

190 Clearly, ϵ is a complex quantity and can be denoted by

$$191 \quad \epsilon = \epsilon' - j\epsilon'' \quad (4.7)$$

192 where ϵ' and ϵ'' are the real and imaginary parts of the permittivity and can be obtained by equating the real and
 193 imaginary parts of equations (4.5) and (4.7). The relationship between electrical conductivity σ and ϵ'' is derived
 194 from Maxwell's equations and it is

$$195 \quad \sigma = \omega\epsilon'' \quad (4.8)$$

196 The velocity of bound charge motion $v = dx/dt$ can be obtained from equation (4.2), such that

$$197 \quad v(\omega) = [(q/m)E]/[\nu - j(\omega_s^2 - \omega^2)/\omega] \quad (4.9)$$

198 The finite velocity of charge motion in the material media indicates that the particle cannot respond
199 instantaneously to a suddenly applied electric field. This time-delay phenomenon gives rise to a frequency-
200 dependent behaviour of charge displacement leading to changes in permittivity with frequency or the relaxation
201 mechanism of interaction of electromagnetic radiation with biological systems. It is noteworthy that the same
202 conclusions are reached by performing the inverse Fourier transforms of equations (4.5) and (4.9) and examining
203 the phenomenon in the time domain. Note that the dependence of permittivity on source and characteristic
204 frequencies ω , ω_p and ω_s suggests that the charge displacement and motion given by equations (4.2) and (4.9),
205 respectively, can also be resonant in nature.

206 Proteins also contain charged groups which are located at sites specific to the atomic arrangements of
207 the molecule. Similarly to isolated ions, these charged groups are bound to water molecules, therefore dielectric
208 properties of biological tissues (at RF) strongly depend on and vary with water content.

209 **4.4 Non-thermal effects (Possible low level interaction mechanisms)**

210 A biophysical mechanism can be specified as non-thermal if the interaction of the RF EMF with living
211 material leads to specific effects other than through heating (Glaser, 2005). Experimentally observed effects are
212 often termed non-thermal when they are not accompanied by a predictable or measurable temperature increase. It
213 is however difficult to ensure that small localized temperature increases, in a cell culture for example, have not
214 occurred during RF exposure.

215 Non-thermal effects (or effects associated with a negligible increase in temperature) can be defined as
216 biological effects that occur with body temperature changes that are either below 1°C, below what is measurable,
217 or in the range of thermal noise. Several arguments have been presented against the plausibility of a non-thermal
218 mechanism existing by which RF radiation could affect physiological changes. These include:

- 219 • Damping effects of the water surrounding biological structures is too strong to allow resonances to exist
220 at radiofrequencies (Adair, 2002);
- 221 • The relaxation time – the time for a molecule to return from an excited state to equilibrium – for
222 excitations produced by RF fields (e.g. vibrations in molecules), is similar to the relaxation time for
223 thermal noise, and shorter than the lifetime of the absorption and transfer of energy into resonant modes
224 of oscillating elements in biological systems (Adair, 2003);
- 225 • The perturbation of the biological structure induced by the applied field must be greater than the effects
226 of random thermal motion and the effects of other dissipative forces, such as viscous damping by the
227 surrounding medium (Foster, 2000). Random thermal motion of charged components in biological
228 systems (i.e. thermal noise) creates random fluctuating EMFs.

229 Based on these arguments, Adair (2003) has concluded that it is unlikely that RF radiation with a power
230 density of less than 10 mW/cm² (100 W/m²) could have a significant effect on biological processes by non-thermal
231 mechanisms. Therefore, it is theoretically implausible for physiological effects (except for reactions mediated by
232 free radical pairs) to be induced at exposure intensities that do not cause an increase in tissue temperature (Adair,
233 2002; 2003; Foster, 2000; Sheppard, Swicord & Balzano, 2008).

234 Sheppard et al. (2008) have also evaluated several potential mechanisms of interaction of RF radiation
235 with biological systems and concluded that, other than heating and possible effects on reactions mediated by free
236 radical pairs, RF field strengths in excess of system noise (collisions among various molecular oscillators
237 generated largely by thermal agitation) could not alter physiological activities without also causing detectable
238 tissue heating. For example, in order for RF electric fields to induce small changes in protein structure that would
239 affect binding of substrates or ligands to enzymes or receptor proteins, extremely high field strengths would be
240 required ($\sim 10^9$ V/m). Sheppard et al. (2008) have addressed mechanistic considerations of interactions as
241 explained in the following sections.

242 **4.4.1 Endogenous electric fields**

243 Endogenous (internally arising) electric fields include quasi-static fields that guide and orientate cells
244 during processes such as embryonic development and wound healing. Other physiological processes give rise to
245 fields that have varying amplitude and phase in response to ever-changing physiological surroundings (Sheppard,
246 Swicord & Balzano, 2008). Fields generated by muscles (including the heart) and the nervous system (including

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247 the brain) are summations of discrete electrical pulses from a large number of electrically active cells; therefore
248 they are not harmonically pure. Their spectrum is continuous and generally confined to certain frequency regions
249 up to a few hundred Hz.

250 Endogenous quasi-static fields have strengths in the range 1–200 V/m (Nuccitelli, 1992), while time-
251 varying fields associated with the central, peripheral and autonomic nervous systems are smaller. For example,
252 Hart and Gandhi (1998) found fields associated with cardiac processes to be a few tenths of a volt per meter in the
253 heart tissues and a few millivolts per meter a distance away from the heart. At cellular membrane level and at
254 sufficiently low frequencies, since the membranes have high resistivity and capacitance (nearly constant for all
255 mammalian cells and equal to 1 F/cm^2), high fields can be produced at the two faces of the membrane. Fields
256 inside the cell are small, as long as the frequency of the applied external field is below the membrane relaxation
257 frequency, which in turn depends on the total membrane resistance and capacitance (WHO, 2007). Gabriel et al.
258 (1996) indicate that the membrane relaxation frequency is typically in the hundreds of kilohertz region.

259 Any exogenous (externally arising) electric field must be of the same order as the endogenous fields (or
260 stronger) to affect normal physiological functions or any functions associated with them (Adair, 2003; Sheppard,
261 Swicord & Balzano, 2008). Therefore, applied fields much smaller than naturally occurring fields in the same
262 frequency band, can only produce a small perturbation in an on-going process.

263 Nuccitelli (2003) suggested that a field of 0.1 V/m would not affect a biological system. However,
264 active neuronal circuits studied in vitro in brain slices, respond to extremely low frequency (ELF) electric fields at
265 about this level; while effects on individual neurons required much greater field strength. Furthermore, no
266 mechanisms for inducing changes in cell membrane potential at frequencies above ~10 MHz have been
267 demonstrated.

268 Applying short pulses (~100 μs) of strong electric fields (e.g. 10–100 kV/m) to cell membranes in order
269 to induce transient pores is called electroporation and allows uptake of drugs, DNA, or other membrane-
270 impermeable substances (Foster, 2000; Sheppard, Swicord & Balzano, 2008). These changes occur without
271 causing significant tissue heating and any consequent thermal damage.

272 Endogenous fields have different characteristics at low and high frequencies. The low frequency power
273 density of these signals in humans is dominated by electric fields from the heart, leading to a maximum of 0.4–0.6
274 V/m in the torso and 15 mV/m in the brain. The spectrum of living tissue in the megahertz region and above has
275 the characteristics of a black body at ~ 300 K (see Chapter 2) and shows no sharp peaks in emission or absorption
276 (Sheppard, Swicord & Balzano, 2008).

277 **4.4.2 Specialized sensory systems**

278 Living organisms have electromagnetic field sensing systems that detect heat and light. The signal
279 detecting structures and signal processing by the nervous system utilises environmental signals as stimuli
280 necessary for survival (Adair, Astumian & Weaver, 1998; Torre et al., 1995). Some species detect quasi-static and
281 low frequency electric fields for use in orientation, navigation and defence (Sheppard, Swicord &
282 Balzano, 2008).

283 While specific sensory systems have been shown to exist for low-frequency, infrared and visible
284 radiation, there is no evidence of comparable RF-sensitive receptors in biological systems. Sheppard et al. (2008)
285 argue that the reason for this is that the RF spectrum lacks natural significant coherent sources and was essentially
286 featureless until the advent of modern man-made sources of RF EMF. In other words, the biological systems did
287 not develop ways to detect RF because this would not have given them any survival advantages.

288 **4.4.3 Effects of weak RF fields**

289 Weak RF fields that do not cause heating would be likely to require frequency-dependent resonant
290 absorption or multiple-photon absorption to induce an amplified signal strong enough to overcome intrinsic
291 molecular thermal noise (Sheppard, Swicord & Balzano, 2008). This is because the photon energy of RF radiation
292 is much smaller than molecular thermal energy at body temperature.

293 Also, biological systems appear to absorb RF signals like a broadband receiver rather than eliciting line
294 spectra characteristic of resonant vibrational motion (Prohofsky, 2004; Sheppard, Swicord & Balzano, 2008). In
295 addition, RF electric field strengths of up to 200 V/m cannot transfer sufficient energy to biological molecules to

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296 alter biological activities or affect thermal noise (kT) fluctuations, such as the opening of voltage-gated ion
297 channels, spatial arrangements of membrane-associated ions, collision rates of charged ligands with proteins, or
298 enzyme reaction kinetics (Adair, 2002; 2003; Sheppard, Swicord & Balzano, 2008).

299 Adair (2002) suggested that, while coupling of RF EMF to biological systems may exhibit resonance
300 behaviour, strong damping of the vibrational motion by interactions with the aqueous environment prevents the
301 absorption of sufficient energy to induce a biological effect. To significantly affect a biological system, the
302 response from the RF signal must be comparable to or greater than the effect of thermal noise (Adair, 2003).

303 A biophysical theory of how low-intensity RF-EMF exposures might affect physiological functions
304 involves the alteration of ligand binding to hydrophobic sites in receptor proteins (Chiabrera et al., 2000).
305 Collisions of the ligand ion in the hydrophobic region of the receptor protein would result in loss of its vibrational
306 energy. In order for RF exposures to affect the binding probability of an ion ligand with a membrane protein
307 receptor, basal metabolic energy would have to amplify the effect of the RF field by maintaining the cell in
308 thermodynamic non-equilibrium. Otherwise, the low-intensity exposure would be negligible compared with
309 thermal noise. Other elements of the model that was used to evaluate the effects of low-intensity RF exposures on
310 ligand binding are the extremely fast (“instantaneous”) rearrangement of atoms in the hydrophobic core of protein
311 by the ligand ion, the fact that the endogenous field at the protein boundaries is large enough to exclude water
312 molecules from the hydrophobic core, and that the ion-collision frequency near the hydrophobic binding site is
313 much less than it is in water. Chiabrera et al. (2000) recognised that thermal noise must be taken into account
314 when evaluating potential biological effects of RF exposures.

315 **4.4.4 Effects of EMF on specific sites**

316 RF EMF may be directed to specific sites of a biological structure, leading to local areas of enhanced
317 field strength. However, the smallest focal spot of concentrated energy would have a radius of the order of a
318 wavelength, which is much larger than most cells (e.g. at 300 GHz, $\lambda = 1$ mm). Thus, on a cellular basis, RF-
319 energy absorption is very small.

320 Fröhlich (1968) has suggested that incident RF energy may be captured by a large group of oscillating
321 dipoles and integrated into a single mode of coherent vibrational energy. For this to occur and produce a coherent
322 response, Sheppard et al. (2008) suggested that the energy stored in the coupled oscillators would need to be
323 comparable to thermal energy and protected from damping by water or other molecules. In addition, energy and
324 thermal diffusion prevent the formation of significant temperature differences at the cellular and subcellular levels.

325 **4.4.5 Non-equilibrium and non-linearity**

326 Since living systems are not in thermal equilibrium, mechanistic theories on interactions between RF
327 EMF and biological tissues must consider the non-equilibrium and nonlinearity of these systems. In principle, non-
328 linear processes such as rectification can transduce RF signals modulated at low frequency into the frequency
329 range where physiological systems operate (Sheppard, Swicord & Balzano, 2008). The theories on the potential
330 effects on biological systems of RF energy at low field strengths must account for the facts that biological systems
331 do not exist at equilibrium, that the dynamic nature of these systems is controlled by enzyme-mediated reactions,
332 and that primary effects may be amplified by nonlinear biological processes (Georgiou, 2010).

333 Binhi and Rubin (2007) suggested that biochemical effects may be induced by weak EMFs in targeted
334 systems that are in non-equilibrium states in which the time to transition from an intermediate metastable state to a
335 final active or inactive state may be less than the thermalization time of the induced field.

336 Prohofsky (2004) has suggested that protein conformation might be affected by RF radiation if
337 amplitudes of specific vibrational modes are altered. However, only intermolecular vibrational modes of proteins
338 and the surrounding tissue occur at radio frequencies and intra-molecular resonant vibrational modes only exist
339 above several hundred GHz. Prohofsky (2004) concluded that the biological effects of RF radiation on
340 macromolecules (proteins and DNA) can only be due to temperature changes because the absorbed energy
341 associated with inter-molecular vibrations is too rapidly converted to heat; i.e. coupling of RF EMFs to the
342 surrounding water (damping) occurs before the energy can be transferred to intra-molecular resonant modes.
343 However, a non-thermal effect might still exist if there were a very strong energy coupling between the inter-
344 molecular and intra-molecular modes and if the effect of water can be isolated. Thus, exceptions to the above-
345 mentioned considerations occur for proteins such as myoglobin or haemoglobin, in which the haem group can
346 oscillate in the protein pocket at lower frequencies. A frequency of 184 GHz is the lowest mode in myoglobin.

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347 **4.4.6 Demodulation**

348 Nonlinearity of certain materials can lead to generation of harmonic frequencies and demodulation of
349 signals (Sheppard, Swicord & Balzano, 2008). The possibility that biological tissue can demodulate an RF signal
350 through the non-linear conversion of RF energy and thereby generating a signal within the tissue at the modulation
351 frequency has been studied (Foster & Repacholi, 2004). Generally, RF signals are modulated at low frequencies to
352 which neurons and neuronal networks such as those in the central nervous system (CNS) are particularly sensitive,
353 and so even weak demodulation could be biologically significant. Ionic conduction through membrane ion
354 channels results in demodulation but only at radio frequencies below about 10–20 MHz (Pickard & Barsoum,
355 1981; Pickard & Moros, 2001).

356 Challis (2005) suggested that demodulation of higher frequency pulsed RF signals (e.g. 900/1800 MHz
357 GSM mobile phone signals pulsed at 217 Hz) might produce low-frequency electric fields in tissues. This would
358 require there to be a nonlinear response in the biological sample. Except for the case of an incident flux of RF
359 energy in the form of extremely high field strength pulses that causes mechanical vibrations, most oscillators in a
360 biological system respond linearly to the incident low energy photons in the RF spectrum. The dispersion of RF
361 energy into random molecular motion energy occurs without generating harmonics of the incident signal in the
362 energy spectrum of re-radiated photons by the exposed material.

363 Balzano and Sheppard (2003) considered the possibility that demodulation of high-frequency incident
364 RF signals might arise from nonlinear interactions with biochemically induced transient oscillators in living tissues
365 e.g. the uncoupled electrons of free radicals. If this were to occur, then the spectrum of RF emission energy
366 emitted from the exposed tissue would be altered, producing a second harmonic that would show up as a spectral
367 line at twice the frequency of the incident signal.

368 Sensitive instruments have been developed to detect the presence of frequency-doubling signals
369 produced by nonlinear interactions between amplitude-modulated RF signals and molecular oscillators vibrating in
370 unison in living cells. These are based on exposing biological samples in cavities designed to be resonant both at
371 the fundamental frequency of an imposed signal and its second harmonic (Balzano, 2003)

372 Balzano et al. (2008) constructed a doubly resonant cylindrical microwave cavity and showed that it
373 was able to detect frequency doubling arising from the nonlinearity of an exposed microscopic Schottky diode test
374 structure. Cells with a diode-like nonlinearity could demodulate a modulated RF carrier wave and generate low
375 frequency signals in an exposed biological preparation. However, the results showed that demodulated field
376 strengths are usually very weak. Specifically, the ELF electric field detected by a nonlinear material from an
377 incident ELF amplitude-modulated RF electric field of 100 V/m would be no more than approximately 3×10^{-3}
378 V/m in the ELF band (Balzano et al., 2008). Therefore, the voltage developed across a membrane with a thickness
379 of 10^{-8} m can be expected to be more than 3×10^{-11} V which is approximately 10^7 times smaller than the low
380 frequency membrane voltage noise that limits physiologically significant events in excitable cells (Billimoria et
381 al., 2006; Jacobson et al., 2005; Kole, Hallermann & Stuart, 2006). In summary, the demodulated ELF signal that
382 may exist across the membrane would be irretrievably lost in membrane noise (Sheppard, Swicord & Balzano,
383 2008).

384 Kowalczyk et al. (2010) used such a doubly resonant cavity to expose more than ten different types of
385 cellular and tissue samples (normal and cancerous) to continuous wave fields at around ~880–890 MHz. The input
386 powers were 0.1 or 1 mW, and SAR values were approximately 11 W/kg for cells and 2.5 W/kg for tissues at the
387 higher of these powers. The authors found no evidence of nonlinear energy conversion to twice the frequency of
388 the incident signal.

389 **4.4.7 Enhanced attraction between cells for pearl-chain formation**

390 The pearl-chain effect occurs when molecules and cells move under the influence of RF electric fields
391 and rearrange to form chains along the direction of the field (Schwan, 1982; Sher, Kresch & Schwan, 1970;
392 Takashima & Schwan, 1985). The effect has been observed with RF fields of about 125 V/m and at frequencies up
393 to about 100 MHz..

394 Pearl-chains have been formed with biological materials such as erythrocytes or bacterial suspensions.
395 Under the influence of RF electric fields, electrical charges tend to accumulate on opposite cell surfaces to form
396 induced dipoles, whose orientation changes with oscillations of the field. A dipole–dipole attraction occurs in the
397 process. The attractive forces between the dipoles are enhanced when the cells are in close proximity to each other.

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398 The dipoles then align in the direction of the applied electric field and form chains of many cells or molecules.
399 These chains are mostly single-stranded, although it is possible to form multi-stranded chains.

400 At frequencies up to about 100 MHz, the precise threshold electric field strength needed to produce the
401 pearl-chain effect depends on frequency, cell or particle size, and pulsing parameters of the applied field. At higher
402 frequencies, the induced dipoles have insufficient time to follow the oscillating field to change their directions.

403 Both pulsed (single or multiple pulses) and continuous wave (CW) fields are known to produce the
404 pearl-chain effect, with a time constant that appears to be proportional to $1/E^2$, where E is the field strength. A
405 minimum amount of energy proportional to τE^2 , where τ and E are the minimum pulse width and threshold field
406 strength respectively, is required to overcome the Brownian forces associated with random motion. The minimum
407 average field strength required for pulsed fields to produce pearl chains is equal to the minimum average field
408 strength for CW fields, suggesting that pulsed fields are no more effective than CW fields in inducing the pearl-
409 chain effect. On the basis that the pearl-chain effect can be produced by a single pulse without a significant
410 temperature rise, the pearl-chain effect is regarded as being caused by forces induced by RF electric fields, not by
411 a biologically significant temperature elevation.

412 While these effects have been observed using in vitro preparations of cell suspensions, they seem
413 unlikely to occur in living systems. In principle, blood could be regarded as such a cellular suspension, but its
414 motion is in irregular directions in organs and unlikely to become aligned with the field over appreciable volumes,
415 except perhaps in long straight vessels.

416 **4.4.8 Magnetite**

417 Magnetite (Fe_3O_4), found in magnetosomes that are present in the human body, including brain tissue, is
418 a strong absorber of RF radiation between 500 MHz and 10 GHz (Kirschvink, 1996). Low-frequency magnetic
419 fields might also produce biological effects if they induce ferromagnetic resonance in tissues that contain high
420 concentrations of magnetite (Challis, 2005). However, magnetite's concentration is very low (5–100 ppb) in
421 human tissues and the resultant heating should be biologically unimportant at localized SARs below guideline
422 levels (Adair, 1996; Kirschvink, 1996; Pickard & Moros, 2001).

423 **4.4.9 Electron tunnelling**

424 Electron tunnelling, the transfer of electrons through biochemical pathways, is a vital element of energy
425 transduction pathways in living cells (Winkler et al., 1999). The electron donor and acceptor are usually weakly
426 coupled in biological systems (Balabin & Onuchic, 2000). Electron tunnelling between donor and acceptor sites in
427 proteins can proceed via a few or multiple transfer pathways. Electron transfer rates are high when the pathways
428 are simple or there is constructive interference amongst multiple paths. However, the rates are significantly lower
429 for destructively interfering pathways (Sheppard, Swicord & Balzano, 2008). Electron tunnelling in proteins is
430 also relatively slow at very long molecular distances (Winkler et al., 1999).

431 Electron tunnelling may occur for a group of similar tunnelling pathways that constitute a tube.
432 Tunnelling via tubes can be strongly influenced by changes in the spatial relationship of donor and acceptor sites
433 according to protein conformation and nuclear dynamics. However, none of these are perturbed by the low photon
434 energy of RF signals (Sheppard, Swicord & Balzano, 2008).

435 If the RF energy absorption is intense, electron tunnelling can be influenced by a temperature increase
436 within complex molecular structures, e.g., proteins. The tunnelling rates are increased by atomic and molecular
437 agitation, opening tubes with constructively interfering pathways or relieving the destructive interference in other
438 tubes. However, very high levels of SAR (>100 W/kg) are required to affect protein conformation (Webb, 1980;
439 Bohr and Bohr, 2000) (Bohr & Bohr, 2000a; b; Webb, 1980).

440 **4.4.10 Radical pair mechanism**

441 Free radicals, which are highly reactive and short-lived molecules or ions with unpaired electrons, are
442 formed when radical pairs dissociate. Scission of a covalent bond in a biological molecule results in the formation
443 of a radical pair, usually as an intermediate stage in some metabolic reaction. If the radical pair lives long enough,
444 a magnetic field can affect the probability of radical recombination and thereby change the reaction yield.

445 Low-intensity magnetic fields may increase the concentration of free radicals by altering the
446 recombination of short-lived radical pairs with antiparallel spins (Challis, 2005; Georgiou, 2010). The expected
447 increase in radical concentration is 30% or less (Timmel et al., 1998). The extent to which this increase can
448 produce oxidative stress-induced tissue damage (e.g. membrane-lipid peroxidation or DNA damage) is not known.
449 Radicals are involved in intracellular signal transduction as part of normal cellular physiology (Finkel, 2003).
450 Therefore, even small effects on radical concentration could potentially affect multiple biological functions.

451 To affect DNA recombination and thus the repair of damage caused by radicals, external magnetic
452 fields must act over the times that the radical pairs dissociate ($>10^{-9}$ s). Based on this, Adair (2003) concludes that
453 the effect of RF fields on free-radical concentrations would likely be limited to frequencies of about 10 MHz or
454 less.

455 Ritz et al (2009) identified a radical pair with a long life time involved in the birds' magnetic compass.
456 The authors studied the effect of external magnetic fields at several frequencies ranging from 0.01 MHz to 7 MHz,
457 with an intensity of 470–480 nT, on robins' behaviour and demonstrated that fields of around 0.6 MHz and above
458 caused them to be disoriented consistently. They concluded that an intense resonance at a Larmor frequency of
459 1.315 MHz (with a 46 μ T static geomagnetic field) is expected for a radical pair in which a radical has a
460 magnetically isolated spin. The strong resonance at a frequency proportional to the intensity of the static field
461 appears to arise from the interaction of the unpaired electron with the external magnetic field to produce a unique
462 energy-level splitting (Zeeman interaction).

463 Georgiou (2010) reported some studies that provide evidence for the induction of oxidative stress via
464 the free-radical pair mechanism in biological systems exposed to RF radiation. Some of the reported effects
465 include increased production of reactive oxygen species, enhancement of oxidative stress-related metabolic
466 processes, an increase in DNA single-strand breaks, increased lipid peroxidation, and alterations in the activities of
467 enzymes associated with antioxidative defence. Many of the changes observed in RF-exposed cells were prevented
468 by (pre)treatment with antioxidants.

469 Sheppard et al. (2008) noted that despite the great number of biochemical reactions involving free
470 radicals, there are many restrictive conditions that would make the effect of RF magnetic fields very unlikely in
471 most systems. These include the frequency constraints based on hyperfine coupling strength, radical pair
472 interactions restricted by the necessity for creation of spin-correlated radical pairs that remain in close proximity,
473 radical lifetimes long enough to be affected by an oscillatory magnetic field, relaxation processes slow enough to
474 allow adequate radical lifetime, and static magnetic fields of appropriate field strength.

475 **4.4.11 Biochemical studies**

476 Biochemical studies are usually carried out on cell-free systems such as proteins, membranes and
477 liposomes in order to acquire information on the validity of hypotheses made at the physical or biophysical level
478 and about the way RF exposure might trigger biological effects, possibly leading to health effects.

479 *4.4.11.1 Biological macromolecules*

480 Some studies have addressed the effects of RF exposure on the structure and function of biological
481 macromolecules such as proteins or DNA. These studies aim to investigate whether absorption of RF energy by
482 macromolecules could modify their structure and/or perhaps their behaviour (Fröhlich, 1968).

483 Bohr and Bohr (2000a) performed a series of experiments on globular proteins, particularly
484 β -lactoglobulin. They exposed protein solutions to RF signals for 5 s in a microwave oven at 2.45 GHz and 800
485 W, causing a $\sim 0.3^{\circ}\text{C}$ temperature increase. Using optical rotational dispersion, the authors showed that exposure
486 accelerated conformational changes of the protein. In a second paper Bohr and Bohr (2000b) reported an
487 enhancement of folding and denaturation of the proteins. These observations were interpreted as evidence of
488 coherent RF excitation of vibrational or torsional modes leading to altered conformation of the protein molecules.
489 However, these results did not consider the difficulty of direct excitation of vibrational modes by RF nor the
490 effects of damping (Adair, 2002; Challis, 2005).

491 Some studies have compared the effect of RF exposure on activity of a thermophilic β -galactosidase
492 with that of conventional heating (Bismuto et al., 2003; La Cara et al., 1999; Mancinelli et al., 2004) and failed to
493 report any changes on structural organization of myoglobin molecule, its internal dynamics and CO (carbon
494 monoxide) binding affinity. The small-amplitude effects of protein misfolding observed by Mancinelli et al.

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495 (2004) are prone to artefacts caused by small variations in the temperature control of the samples and these results
496 are yet to be confirmed (ICNIRP, 2009).

497 Specific effects on a solution of green fluorescent protein exposed at 8.5 GHz were reported by Coptly et
498 al. (2006). Samples were either exposed to RF or heated using resistive heating with maximum RF power
499 corresponding to a calculated SAR of 4 kW/kg and ΔT of 3°C. In both cases, heating produced a decrease in the
500 protein fluorescence intensity and the spectrum became red shifted. For a similar temperature rise, the alteration of
501 fluorescence was larger in the RF-exposed samples, which was interpreted as evidence of a specific non-thermal
502 effect of RF exposure. However, the theoretical and experimental determination of ΔT under RF exposure is very
503 uncertain for any conclusion regarding non-thermal effects (ICNIRP, 2009).

504 There have been some studies on isolated DNA in solution, reporting a frequency-specific absorption in
505 DNA from plasmids or DNA breakage due to RF exposure in solution (Edwards et al., 1984; 1985; Sagripanti &
506 Swicord, 1986; Swicord & Davis, 1982). However, follow-up studies failed to confirm the early results (Foster,
507 Epstein & Gealt, 1987; Gabriel et al., 1987). DNA breakage was most likely to have been the result of free radical
508 formation due to the use of copper electrodes and hence the presence of copper ions in solution, but not the result
509 of a direct action of RF absorption (Sagripanti, Swicord & Davis, 1987). Further theoretical calculations by Foster
510 and Baish (2000), Adair (2002) and Prohovsky (2004) support the view that viscous damping would be sufficient
511 to make any 'resonant' behavior of DNA molecules in solution very unlikely.

512 4.4.11.2 *Liposomes and membranes*

513 Liposomes are artificial phospholipid vesicles, constructed in the laboratory, which have often been
514 used as models for studies of membrane properties. Early work by Liburdy and Magin (1985) reported an
515 enhanced release of drugs trapped in the liposomes under exposure at 2.45 GHz with an SAR of 60 W/kg. The
516 effect occurred at temperatures below the membrane phase transition temperature of 41°C.

517 Ramundo-Orlando et al. (2004) exposed liposomes entrapping glycoenzyme ascorbate oxidase to
518 2.45 GHz, and SAR levels of up to 5.6 W/kg. Exposure at the maximum SAR level reduced enzyme activity,
519 although the conformation of the enzyme was not affected. The authors suggested that RF interactions with the
520 oligosaccharide chains of the enzyme were critical in eliciting this effect. Further work by the same group at
521 130 GHz using the carbonic anhydrase enzyme led to increased liposome permeability under pulsed exposure, but
522 only when modulation was at 7 Hz and not 5 or 10 Hz (Ramundo-Orlando et al., 2007).

523 4.5 **Summary**

524 A review of established and proposed mechanisms reported in literature suggests that the most
525 important mechanism of interaction between RF and biological systems is the heating of tissues by dielectric and
526 resistive loss.

527 Other mechanisms have been hypothetically proposed to understand experimental observations that
528 could not be explained by thermal mechanisms. However, as Sheppard et al. (2008) demonstrated, detailed
529 analysis of proposed molecular mechanisms showed that over-damping by water imposes strict limitations on
530 vibrational or rotational modes and precludes a direct RF pumping of such modes.

531 Some of the proposed mechanisms involve energy levels that are much smaller than the thermal
532 background and, even considering various circumstances for amplification, could not overcome the influence of
533 thermal noise (Sheppard, Swicord & Balzano, 2008). Those mechanisms governed by quantum mechanics are
534 coupled to the thermal noise and subject to the same limitation. Therefore, Sheppard et al. (2008) concluded that
535 there is no mechanistic path whereby RF energy can be directly imparted to selected oscillatory modes of the
536 biomolecular system.

537 Some proposed mechanisms involve the existence of nonlinear processes in which some of the energy
538 in an RF signal is transferred into lower frequency bands where there are metabolic oscillations, leading to
539 interference with those oscillations. Demodulation of amplitude modulated, including pulsed signals is possible
540 below a radio frequency of about 10 MHz so non-thermal effects could then in principle occur at a lower
541 modulation frequency. However, even without considering demodulation efficiency, the maximum theoretical
542 power that can be demodulated is extremely small (Sheppard, Swicord & Balzano, 2008).

543 RF energy below thermal levels can affect radical pair-dependent chemistry with possible biological
544 consequences. This phenomenon reflects the quantum mechanical nature of energy level splitting via the hyperfine
545 interaction for certain nuclei of a radical pair. However, magnetic resonance in the gigahertz regime requires a
546 nucleus with an exceptionally large hyperfine coupling constant, indicating that effects on free radical reaction
547 rates are unlikely to be a general feature of biochemistry (Sheppard, Swicord & Balzano, 2008).

548 Overall, the search for non-thermal effects of RF on biological macromolecules such as proteins and
549 DNA has not generated good evidence to suggest that such effects occur. Overall, there is limited evidence to date
550 that non-thermal RF effects occur in model liposomes although the biological significance of such effects is not
551 clear.

552

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